

Neuromodulation of Neural Oscillations in Health and Disease

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Simple Summary: Over the past few decades, advances in electroencephalography (EEG) recordings and brain stimulation has permitted an unprecedented view of how specific brain structures communicate as well as organize complex cognitive functions. Specifically, neurotransmitters (including norepinephrine, acetylcholine, and dopamine) have all been shown to have an impact on neural oscillations throughout the brain, linking them to changes in cognitive functions such as memory, attention, and executive function. While these interactions are still widely unexplored, their appearance in neurological disorders through cross-frequency coupling (CFC) brings light to the vital role they play in orchestrating healthy brain function. This brief review serves to highlight the important role each neuromodulatory system plays in changing widespread neural networks, emphasizing their involvement in health and disease to help inform more translational brain stimulation technologies.

Abstract: Using EEG and local field potentials (LFPs) as an index of large-scale neural activities, research has been able to associate neural oscillations in different frequency bands with markers of cognitive functions, goal-directed behavior, and various neurological disorders. While this gives us a glimpse into how neurons communicate throughout the brain, the causality of these synchronized network activities remains poorly understood. Moreover, the effect of the major neuromodulatory systems (e.g., noradrenergic, cholinergic, and dopaminergic) on brain oscillations has drawn much attention. More recent studies have suggested that cross-frequency coupling (CFC) is heavily responsible for mediating network-wide communication across subcortical and cortical brain structures, implicating the importance of neurotransmitters in shaping coordinated actions. By bringing to light the role each neuromodulatory system plays in regulating brain-wide neural oscillations, we hope to paint a clearer picture of the pivotal role neural oscillations play in a variety of cognitive functions and neurological disorders, and how neuromodulation techniques can be optimized as a means of controlling neural network dynamics. The aim of this review is to showcase the important role that neuromodulatory systems play in large-scale neural network dynamics, informing future studies to pay close attention to their involvement in specific features of neural oscillations and associated behaviors.

Keywords: neuromodulation; EEG; noradrenergic system; cholinergic system; dopaminergic system; pupil-linked arousal; neural oscillations; cross-frequency coupling; vagus nerve stimulation; neurological disorders

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1. Introduction

Neural oscillations are thought to be an essential driver of interaction, communication, and information transmission throughout the brain [1–3]. Evolution has maximized the role these oscillations play in regulating and controlling neuronal functions, driving the synchronization of widespread neural networks in the brain. The EEG provides the most popular non-invasive methods to record neural oscillations, summing the local field potentials of thousands of neurons in cortical structures [4,5]. Not only does this tool

characterize the “global” brain state as a time series of voltage potentials, but also allows researchers to analyze these oscillatory waveforms through frequency domain analysis. Studies have suggested that distinct EEG frequency bands (Delta, Theta, Alpha, Beta, Gamma) are generated from unique neural populations across a variety of brain regions [6]. This characterization shows the ability of different brain structures to generate specific neural oscillatory patterns, permitting synchronization and frequency coupling [6]. Assessing the effect of oscillatory changes both globally and locally throughout the brain can uncover the important and/or causal role of various neuromodulatory processes [7]. Cross-frequency coupling (CFC), resulting from coupling between various neural circuits and/or different types of neurons through chemical or electrical synapses, has recently become a more prominent topic [8,9]. Components of CFC such as phase–phase and phase–amplitude coupling have been shown to have a large influence on cognitive processes including attention, learning, and short- and long-term memory [10,11]. Additionally, the phase–amplitude synchronization of these high and low frequency bands plays a prominent role in facilitating neural communication and neural plasticity [12–15]. Neurological diseases and conditions can often be associated with an abnormal oscillatory desynchronization or type of CFC, unveiling the importance that synchronized neural networks have in carrying out normal brain function [16]. While largely misunderstood, this network-wide communication seems to be an instrumental part of the coordination and regulation of cognitive abilities. More recently, neural stimulation techniques such as vagus nerve stimulation (VNS) and deep brain stimulation (DBS) have been incorporating types of CFC analysis to better understand the effects of phase-coupled neuromodulation [17–20]. By leveraging the causal effects of neuromodulation on cognitive functions, these tools are focusing on real-time oscillation analysis to optimize the effectiveness of stimulation across brain regions [21].

The neuromodulatory systems, including the noradrenergic, cholinergic, and dopaminergic systems, play a pivotal role in the regulation and synchronization of neural oscillations. These systems provide direct axonal projections to most structures of the brain, regulating various brain functions through the release of neurotransmitters [22–25] (Figure 1a). Specifically, norepinephrine, acetylcholine, and dopamine are all implicated in the formation of complex decision making and executive functions [26]. Through their activation and inhibition, each neuromodulatory system has been seen to change the oscillatory behavior of widespread neural networks, implicating changes in cortical structures as well as in different frequency bands [27]. Recently, more work has been conducted that focuses on how cross-frequency coupling can be affected by neuromodulation, with phasic or tonic neurotransmitter release causing synchronization or desynchronization in EEG waveform features such as power, amplitude, phase and frequency [28]. These experiments are often using optogenetic manipulation in conjunction with LFP recordings, providing insights into how particular neuromodulatory centers can have a profound effect on neural oscillations, even in indirectly coupled brain regions. Another non-invasive biometric measure, pupil size, has also been implicated in having a key role in indexing neuromodulation, potentially serving as a new indirect modality in understanding the widespread effect of different arousal states on neural oscillations and behavior [29–34].

This review will focus on how these three neuromodulatory systems can individually modulate neural oscillations, looking specifically at how their activation can change large-scale neural network synchrony in cognitive functions and neurological disorders. This phenomenon has yet to be fully understood, and looking at how each system’s activation or inhibition affects large scale oscillatory patterns may help uncover the origin and causality of complex behaviors and neurological disorders. These insights will hopefully inform a new direction of research that looks to further investigate how neuromodulation could improve and/or shape brain functions through changing large-scale neural oscillations.

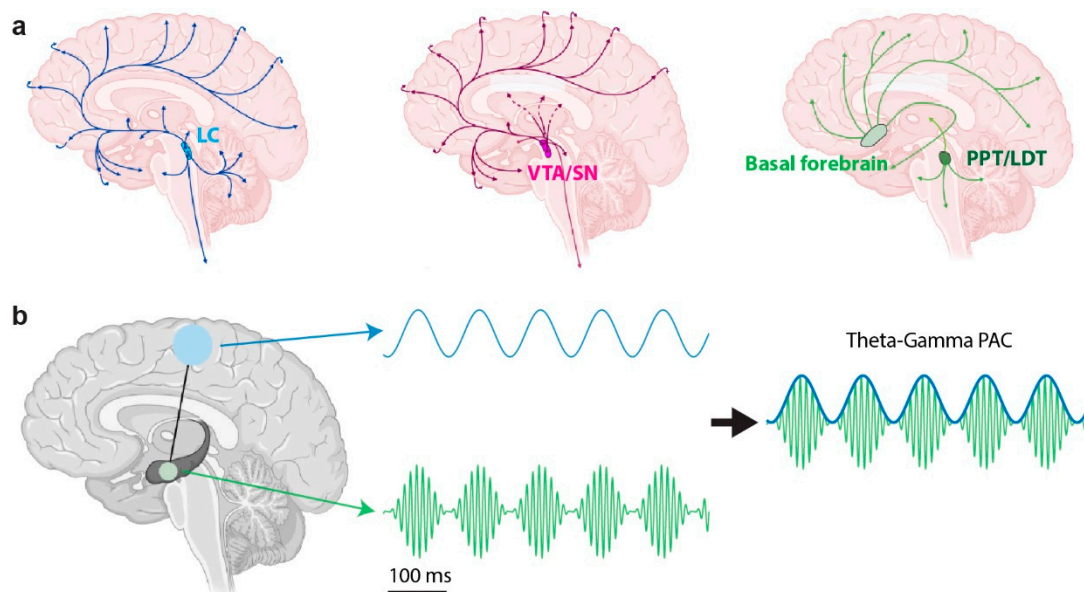


Figure 1. Anatomical representations of each neuromodulatory system and CFC visualization: (a) Anatomical locations and projections of the three neuromodulatory systems: the noradrenergic system (left), dopaminergic system (middle), and cholinergic system (right). (b) A cartoon illustrating theta-gamma phase-amplitude coupling (PAC).

2. Neural Oscillations in Cognition and Neurological Disorders

2.1. Frequency Spectral Analysis

Over the past few decades, an abundance of research has been conducted exploring the role different frequency bands play in representing the neural signals behind cognition, behavior, and neurological disease. These have been characterized by abstracting the frequency spectrum into five unique frequency bands: Delta (1–3 Hz), Theta (4–7 Hz), Alpha (8–12 Hz), Beta (13–30 Hz), and Gamma (30–100 Hz) [35]. While an abundance of studies has implicated each band in a variety of diseases and cognitive functions, generally, slow and high oscillations can be discriminated by specific behavioral outputs. It is important to acknowledge that these oscillations are often shaped by the summations of thousands of neurons in a particular brain region, making these associations highly dependent upon the type of recording device (EEG, ECoG, invasive LFPs) and their signal-to-noise ratio. Even with this in mind, looking at how each frequency band is implicated throughout complex cognitive processes can make it easier to understand how neuromodulation plays a vital role in shaping these neural oscillations.

2.1.1. Healthy Functions

Throughout normal cognition, each oscillatory frequency band has been linked to a specific behavioral correlate or executive function. Generally speaking, delta waves are associated with a wide variety of cognitive functions. Studies have shown that the delta oscillatory phase is correlated with the reaction time of behavior, playing a role in the synchronization of neural populations across multiple brain regions [36]. Additionally, these oscillations are often associated with a resting state, with the slow oscillations (less than 1 Hz) having the ability to trigger thalamically-generated spindles [37]. Theta oscillations are thought to play a vital role in almost every cognitive function and brain region. Studies have uncovered their critical role in learning, memory, and synaptic plasticity. It is also a foundational band for cross-frequency coupling, synchronizing with the gamma band in a phenomenon known as theta-gamma coupling that is heavily studied for its role in working memory [38–41]. The alpha band is also one of the most studied frequency bands, specifically for its clear role in attentional demands and visual perception as well as its easily triggered appearance in stimuli-associated tasks [42–47]. The beta band has

often been associated with motor-induced events [48]. The synchronization of beta frequencies has been an indicator for normal motor system functioning, and it has been shown to play a role in working memory [49,50]. Finally, the gamma band, like the theta band, plays a vital role in memory, as seen in theta–gamma coupled oscillations throughout the hippocampus and cortex [41]. Gamma also has appearances in attention, consciousness, and wakefulness experiments [51–53].

2.1.2. Neurological Diseases

While each frequency band is instrumental to maintain healthy brain function, neurological diseases seem to alter specific oscillatory properties, oftentimes leading to cognitive dysfunction. To begin, abnormal delta oscillations have often been linked to neurological conditions associated with sleep. Specifically, non-rapid eye movement sleep in both Schizophrenia and Alzheimer’s disease has been associated with a depressed delta power when compared to healthy patients [54,55]. Clinically, the theta band has been linked to the maintenance of the healthy human brain. Studies have explored the loss of long-range temporal correlations in theta oscillations in patients with major depressive disorder as well as increased theta event-related synchronization in children diagnosed with ADHD [56,57]. A slowing in spontaneous alpha oscillations has been consistently linked to Alzheimer’s disease. Additionally, likely due to its prominent role in attention, resting alpha power has been shown to be reduced in adults with ADHD [58,59]. Higher frequency oscillations have been largely linked to motor-related conditions such as Parkinson’s tremors. The beta band has been reported to be desynchronized in Parkinson’s patients [48]. The modulation of beta oscillations through deep brain stimulation has enabled new targeted therapeutic treatments, oftentimes reversing these motor symptoms [60]. Gamma oscillations have also appeared in Parkinson’s disease and during dyskinesia [61]. Likewise, aberrant gamma oscillations have appeared in mouse models of Alzheimer’s disease and Fragile X syndrome [62].

2.2. Cross-Frequency Coupling Analysis

While each frequency band seems to dissociate into a wide variety of behaviors or cognitive functions, the synchrony between each band through cross-frequency coupling (CFC) analysis has brought new light to the complexity of neural networks [63]. Recent studies which uncover the coupling between distinct features of neural oscillations such as phase, amplitude, and power between frequency bands have shown its critical importance to network dynamics, learning, and complex behaviors [64–67]. CFC is thought to be integral to the temporal and spatial activation of specific cortical circuits, with one study showing how the magnitude of gamma oscillations are modulated by slower wave rhythms [68]. By pairing each frequency interaction with another (i.e., power–power, phase–phase, phase–frequency, phase–power, and phase–amplitude), each coupled oscillatory feature provides a unique effect on the synchronization of widespread neural networks and changes in cognitive abilities [10,69–73]. For example, observed phenomena such as theta–gamma coupling, known for its role in working memory, is only one of many oscillatory patterns which has linked the important role of band coupling in neural communication, synaptic plasticity, and executive functions [8,10,40,74–77] (Figure 1b).

2.2.1. Healthy Functions

Previous studies show more instances of CFC playing a role in cognitive functions across memory, learning, attention, and decision making. Specifically, alpha–beta phase–amplitude coupling (PAC) has been seen synchronized in the medial prefrontal cortex during decision making, as well as the phase of delta and theta bands, suggesting the role of PAC in feedback coding [78,79]. In decision making, CFC was seen to be highly correlated with rodent behavioral performance, with CFC strength increasing over time in hippocampal regions [65]. PAC was also seen between alpha and gamma bands in response

to visual grating stimuli [80]. Additionally, attention has been seen in a wide variety of CFC phenomena, with purposeful synchronization across brain regions driving the formation of neural ensembles associated with specific tasks. Specifically, spatial attention experiments showcase two unique PAC clusters, with delta–gamma PAC being sensitive to cue direction and theta–alpha and beta–gamma PAC associated with future reaction times [81]. Similarly, delta rhythm synchronization was correlated with the reaction time to anticipatory signals [36]. Alpha and gamma rhythm synchronization has also been seen to contribute to selective attention, with a study showing increased beta and alpha synchronization in rule-directed task behaviors [82]. Delta–theta phase high gamma amplitude coupling, triggered by attentional demands, was seen as a mechanism for sub-second facilitation and coordination in the parietal cortex [83]. Likewise, the implementation of alpha oscillations in visual discrimination highlights the role of CFC in filtering out stimuli distractors [84].

CFC is also thought to be the brain’s method of transferring large amounts of information across distant regions of the brain in an organized fashion, and it has been shown to impact different forms of learning and memory across species. Theta oscillatory synchronization between the prefrontal cortex (PFC) and the amygdala supports communication across interneuron networks and is associated with fear learning [85]. Theta band synchronization of the anterior limbic system was also shown to be associated with long term memory, while alpha band desynchronization of the posterior thalamic system was associated with short-term memory [86]. Additionally, synchronization of the PFC between 3 and 32 Hz aided in short-term memory [87]. The large variety in synchronization patterns found across short- and long-term memory could be a result of the lack of consistency in behavioral tasks and experimental methods used across diverse research groups. Nevertheless, it could also showcase that optimal short- and long-term encoding for unique memory tasks or objects that occurs at very specific CFC patterns [87]. It is important to note that the CFC studies that we just described were characterized throughout healthy brain function and observed during the natural rhythms of wide-spread neural networks. While these oscillatory phenomena have uncovered interesting links to cognitive functions, their causality is still mostly untested. It is possible that CFC is a predictive biomarker of communications between distinct populations of neurons mediating various brain functions and is therefore subject to the influence of neuromodulatory systems.

2.2.2. Neurological Diseases

In playing such a large role in neural physiology, it is no surprise that CFC has also been involved in a variety of neurological diseases. Being most observed in working memory studies, PAC has been associated with diseases including schizophrenia, obsessive–compulsive disorder, Alzheimer’s disease, epilepsy, and Parkinson’s disease. For example, it has been suggested that CFC may play a role in Parkinson’s disease as excessive PAC between beta and gamma bands has been observed in advanced-stage patients with movement deficits [64,88]. Abnormal theta–gamma CFC has been implicated in Alzheimer’s disease, as coupled band synchronization plays a critical role in memory functions [89,90]. Also, theta–phase gamma–amplitude coupling has been seen as a marker of Attention Deficit Hyperactivity disorder (ADHD) in children [91]. Likewise, schizophrenia studies have revealed adverse CFC in healthy patients as they suffer from clear disruptions in theta and gamma oscillations in the temporal lobe and auditory cortex [92,93]. Disease-driven CFC is rooted in the loss of synchronization between frequency bands that are in coordination in a healthy state. These disruptions are often a product of pathological interference, preventing normal brain function due to a lack of synaptic connections or adverse oscillatory rhythms. While CFC’s role in disease is far from understood, future experimental models should make note of CFC changes, as mapping out these oscillatory dynamics could help enable faster diagnosis and/or more targeted brain stimulation treatment.

Cross-frequency coupling has been found in the synchronization across subcortical and cortical regions, orchestrating a bottom-up circuit where deep brain regions such as the hippocampus and neuromodulatory centers such as the locus coeruleus (LC), ventral tegmental area (VTA), and basal forebrain (BF) have direct control of brain-wide neuronal synchrony. However, this phenomenon has yet to be explored in patients, as invasive stimulation and optogenetic/chemogenetic manipulations are challenging to conduct in human studies. Future experiments using animal disease models would be better positioned to look into how specific brain structures modulate CFC to help uncover the causal relationship between cognitive abilities and neural oscillations in health and disease.

3. The Neuromodulatory System's Role in Neural Oscillations

3.1. The Noradrenergic System

The noradrenergic system plays a very important role in the modulation and regulation of norepinephrine (NE) throughout the brain. The center of norepinephrine creation, the locus coeruleus, is a complex nucleus with projections to almost every area of the cortex and subcortical areas [94–97]. Projections directly from the LC paired with directed synchronous and asynchronous firing patterns of LC neurons have been shown to allow for differentiated and targeted norepinephrine signaling throughout the cortex [96,98] (Figure 1a). The LC releases norepinephrine via two patterns: tonic and phasic firing. Phasic activation, which is commonly seen in rapid behavioral adaptation, is often associated with a large-scale reorganization of targeted neural networks [99,100]. Tonic activation, in contrast, seems to have a prominent role in NE release but lacks the desynchronization associated with phasic activation [101]. Differences within and across phasic and tonic stimulation patterns in LC neurons is implicated in various types of behavioral tasks and attentional processing [102]. While studies have not specifically defined the LC's direct role in widespread neural oscillations, experiments incorporating a type of LC modulation in conjunction with electrophysiology recordings have unveiled the LC's ability to control selective neural oscillatory patterns all the way down to specific frequency bands in localized brain regions. Phasic activation of the LC has been shown to result in clear desynchronization in the EEG spectrum [103,104]. For instance, phasic microstimulation of the LC of rats increased the ratio of EEG power in high frequencies (10–100 Hz) to low frequencies (1–10 Hz), desynchronizing cortical EEG [105] (Figure 2a,b). An abundance of work has also been performed focusing on the LC's role in the hippocampus, with the LC-NE system seen driving long-term potentiation (LTP), increasing theta power, and with gamma power being reduced in rats that did not experience LTP [101]. Each type of noradrenergic receptor, $\alpha 1$, $\alpha 2$, and $\beta 1$, has been implicated in theta oscillations and synchronization in the hippocampal formation [106–109]. Furthermore, studies have shown that theta activity was able to be modulated by reboxetine, a norepinephrine reuptake inhibitor [110]. In regard to cortex-specific cognitive functions, phasic release of norepinephrine has been seen to change the EEG spectrum across the mPFC, guiding motor planning, decision making, and sensory processing [111–113]. The activation and inhibition of subcortical regions has led to alpha wave synchronization which facilitates in directing selective attention [114]. The bidirectional communication between the LC and PFC has been seen to act as a level of control over excitatory input into the LC, marked by a high gamma frequency in the PFC [28]. Optogenetic stimulation of the LC in conjunction with neural spike recording in the mPFC saw increased neuronal firing and persistent spikes, present in high amplitude and slow frequency oscillations (delta and theta waves). These increased depolarization events are often associated with an enhancement of synaptic plasticity and memory consolidation [115]. Taken together, these results suggested that the coupling of phase, amplitude, and power between different frequency bands may be directly influenced by NE release from the LC, with specific stimulation parameters being essential for changes in the synchronization of these oscillatory features. Some recent evidence showed that blocking the noradrenergic system through administration of

Clonidine, an alpha-2 agonist, changed the phase–amplitude coupling in cortical EEG in mice (Figure 2c). Future studies which look to selectively adjust the frequency and power of stimulation, directing a graded release of NE, could potentially provide a type of control over neural oscillations or help restore CFC relationships that may have been lost to neurological disorders. Although a substantial amount of research questions remain unanswered about the true relationship between the noradrenergic system and neural oscillations, these previous studies open the door to a clear causal relationship of the role NE plays in shaping cognitive abilities through widespread oscillatory dynamics.

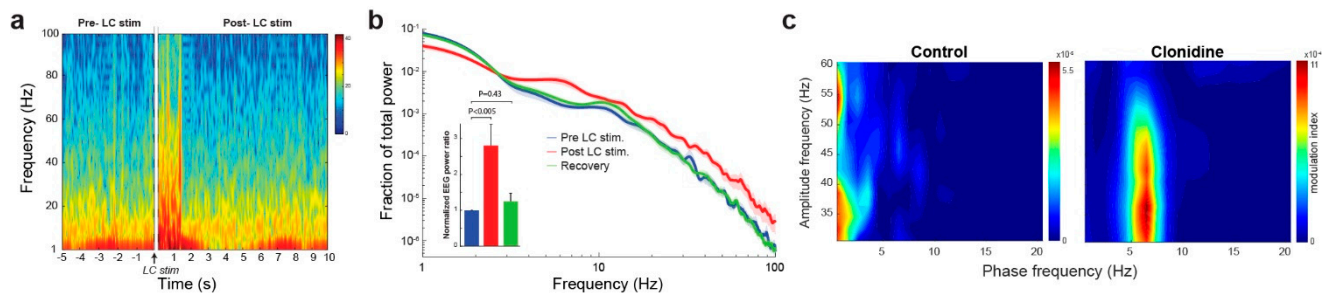


Figure 2. The causal effect of the noradrenergic system on neural oscillations: (a) Spectrogram of cortical EEG around phasic LC stimulation. (b) LC stimulation resulted in a significant increase in EEG power ratio in a high frequency (10–100 Hz) to low frequency (1–10 Hz); blue (pre LC stim), red (post LC stim), green (recovery); adopted from [105]. (c) Manipulation of the noradrenergic system through Clonidine administration, an alpha-2 agonist, altered phase-amplitude coupling of cortical EEG (2 mice, 9 sessions; unpublished data from the authors).

3.2. The Cholinergic System:

The cholinergic system, like the noradrenergic system, also plays a vital role in the control of neural oscillations throughout the brain and cortical structures [116]. Studies have already unveiled its contribution in central nervous system physiology and in various disorders such as dementia, epilepsy, and sleep disorders [117–119]. While acetylcholine (ACh) is more widespread and abundant in the brain than norepinephrine, its neuromodulatory centers, including the pedunculopontine nucleus (PPT), laterodorsal tegmental nucleus (LDT), and basal forebrain (BF), have direct projections to widespread brain regions, allowing the cholinergic system to readily influence the oscillations of neural networks [26]. Retrograde tracing has shown that the cortex has an abundance of projections from the basal forebrain, connecting subcortical structures to the frontal, cingulate, and medial parietal cortex [120,121] (Figure 1a). Specifically, the BF has been seen implicated in the direct regulation of attentional functions through multiple thalamic and cortical projections [122]. Similar to the release of norepinephrine from the LC, acetylcholine is also released throughout the brain in tonic and phasic patterns [123,124]. Studies have shown that tonic acetylcholine release in the prefrontal cortex was coordinated with the hippocampus and was maximal during training on a working memory task. Phasic release, in contrast, only occurred during the memory task and was localized to reward delivery areas, independent of the trial outcome [125]. Although tonic, volume-based ACh release is traditionally viewed to be the driver, recent optogenetic studies suggested a link between phasic ACh activation in the cortex and its causal role in behavioral changes [126].

Like noradrenergic modulation, cholinergic modulation has not specifically been studied in conjunction with cross-frequency coupled phenomena. Instead, studies utilizing widespread neural recordings have looked to uncover the direct role of ACh in behavior and erratic firing patterns associated with neurological disorders. Specific low-frequency oscillations including delta, theta, and alpha have been seen to be suppressed by brainstem cholinergic neurons, while high-frequency bands such as beta and gamma are accompanied by an increased release of ACh in the thalamus and cortex [127]. Supporting

this notion, direct electrical stimulation of the nucleus basalis, the cholinergic nucleus of the basal forebrain, increased the ratio of power in high frequency (10–100 Hz) bands over low frequency (1–10 Hz) bands of cortical LFPs [128] (Figure 3a,b). ACh release in cholinergic neurons has also been seen to discharge at higher rates during cortical activation rather than during slow-wave cortical activity, with thalamo-cortical and brainstem-cortical cholinergic activity initiating theta rhythms and influencing task-specific cortical desynchronization [129]. Cholinergic system modulation is also highly dependent upon input from noradrenergic LC projections. A particular study explored that when norepinephrine was administered into the basal forebrain in sleep-waking rats, it elicited an increase in fast gamma EEG oscillations and a diminution in slow delta activity, but when neurotensin was administered, theta and gamma activity as well as wakefulness were enhanced [130,131]. Furthermore, cholinergic neurons seem to play a vital role in influencing theta oscillations in the hippocampus. Studies have shown a direct relationship between ACh level and theta oscillations in hippocampal neurons, as well as a direct impact on the amplitude of theta waves. This relationship potentially influences neural computations responsible for memory encoding and retrieval [132–135]. Theta–gamma coupling also seems heavily dependent on ACh modulation, with detected cues evoking phasic ACh release as well as neural synchrony between frequency bands in the PFC [126,136]. Likewise, varying regions of high and low ACh signaling lead to the emergence of localized gamma–theta coupling, lending support to the theory of cross communication between brain regions during attentional processes. Specifically, studies have shown that stable gamma-modulated firing occurs in regions with high ACh signaling, implicating its causal role in generating localized theta–gamma rhythms [51].

Oftentimes, ACh-triggered neural synchrony has been explored using activation or inhibition studies through optogenetics and pharmacological agents. Specifically, phasic optogenetic stimulation of the basal forebrain has been seen to modulate the cortical topography of auditory steady state responses, with phase-locked stimulation enhancing the power of cortical responses and increasing their synchronization, as well as changing broadband gamma power [137–139]. Additionally, stimulation applied to the pedunculopontine tegmentum (PPT) significantly increased ACh release that drove desynchronization in the cortical EEG [140]. The relationship between CFC and cholinergic modulation is still a widely unexplored phenomenon, but future research specifically looking at how phasic stimulation parameters could affect different aspects of CFC could provide new insights into how the cholinergic system drives complex cognitive functions through widespread neural oscillations.

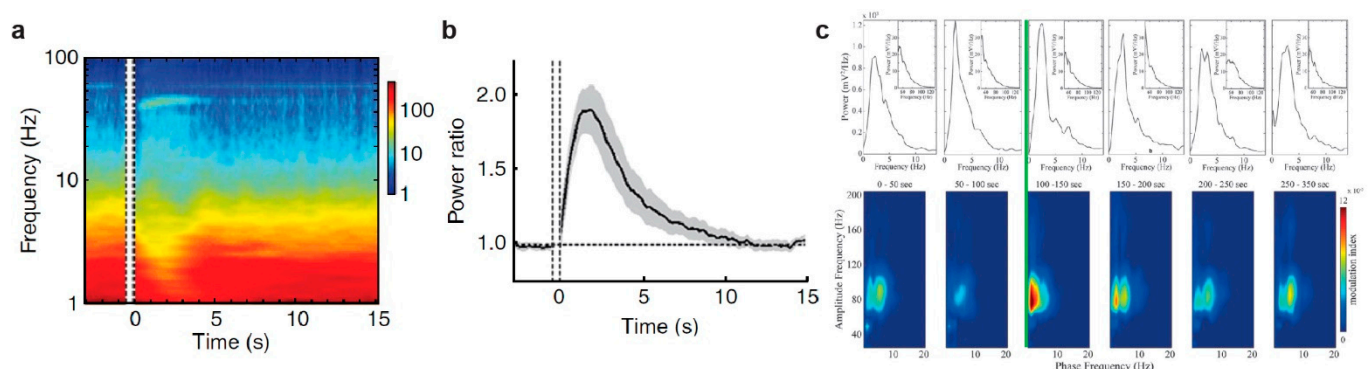


Figure 3. The causal effect of the cholinergic and dopaminergic system on neural oscillations: (a) Spectrogram of cortical LFP around nucleus basalis stimulation. (b) LFP power ratio (power at 10–100 Hz over power at 1–10 Hz) after nucleus basalis stimulation. Adopted from [128] with permission. (c) Dopaminergic receptor activation changed phase–amplitude coupling in mPFC. Adopted from [72].

3.3. The Dopaminergic System:

The dopaminergic system is critically important to the circuitry and control of cognitive functions in the prefrontal cortex [141]. Like the other neuromodulators, dopamine is synthesized in various centers across the brain including the ventral tegmental area (VTA), the hypothalamus, and the substantia nigra (SN) [24] (Figure 1a). Theories have proposed a complex neuronal microcircuit based on the known mechanism of action of dopamine in the PFC, accounting for the diverse role of dopamine in executive functions [142]. The direct projections from the VTA and SN via the mesocortical dopamine pathway are responsible for functions such as working memory, attention, and decision making [143]. Similar to the release of other neuromodulators, tonic and phasic dopamine release have been shown to play different roles in mediating a wide range of brain functions, including learning, motivation, and motor control, possibly through their distinct effects on dopaminergic receptors [144].

The dopaminergic system has been shown to be heavily involved in the shaping of neural oscillations in a variety of cognitive functions and neurological disorders. Studies utilizing electrophysiology recordings have demonstrated the vital role of dopamine activation throughout the brain. Specifically, dopaminergic receptor activation has been linked to weaker alpha and beta oscillations in the PFC, while depletion of dopamine has been linked to increased power in beta oscillations in the cortex and subthalamic nucleus of rats [145,146]. Dopamine also plays a role in working memory, with it facilitating low theta oscillations in the PFC as well as acting as a trigger for latent theta oscillations [147]. Dopaminergic modulation through stimulation and pharmacology has also seen clear effects on cortical state. Dopamine injections in the PFC of anesthetized rats, for example, has been shown to provoke an increase in hippocampal and prefrontal coherence [148]. Stimulation of the VTA has been shown to induce reanimation from anesthesia, with optogenetic stimulation of even a small portion of VTA dopamine neurons being sufficient to induce this transition [149]. Additionally, the emergence of exaggerated beta oscillations in the cortex, a key symptom of Parkinson's disease, is marked by a disruption in dopamine transmission [150]. The synchronization of beta frequencies facilitates normal motor functioning, and dopamine levels largely impact beta synchronization to guide normal function [49]. However, while beta oscillations in cortical and basal ganglia networks are closely coupled to dopamine tone in humans, phase–amplitude coupling appeared not to be directly regulated by dopamine levels. These findings have key implications for Parkinson's patients, and future research is necessary to uncover how these findings can be used to benefit patients' quality of life [151].

The dopaminergic system clearly acts as a controller of large-scale neural oscillations, but its causal relationship with CFC is largely unknown. Looking at dopamine-triggered synchronization studies may help provide context to the role of dopamine in CFC. In Parkinson's patients, chronic dopaminergic transmission interruption promotes excessive cortical beta synchronization that is seen in high coherence between motor and somatosensory cortical activities [152]. Desynchronization in the cortex has been seen through the stimulation of D1 dopamine receptors in behavioral arousal experiments in rats and rabbits [153]. Looking into more specific phase and amplitude phenomena, dopamine release in the mPFC has led to shifts in phase–amplitude coupling from the theta–gamma band to delta–gamma band, giving insight into how dopamine may regulate function in the mPFC [72]. Interestingly, in this study, laser activation of RuBi-Dopa, a light-sensitive caged compound, released dopamine and in turn increased activation of dopamine receptors, resulting in a modulation of CFC throughout LFP recordings in the mPFC but not LFP power in any frequency band [72] (Figure 3c). Furthermore, theta phase coupling between the hippocampus and the mPFC may also be modulated by the dopamine system and could be an underlying mechanism of cognitive dysfunction in depression [154]. Recently, a study showed how phasic dopamine activation in the PFC had a robust influence on coordinated gamma oscillations, initiating a gamma–theta coupling which lasted for

several minutes [155]. Overall, the relationship between cross-frequency coupling and dopamine is still widely unclear. Future research studying how stimulation of dopamine neuromodulatory centers and receptors impact select features of neural oscillations could further help us understand the causality of cognitive functions and neuropsychiatric disorders.

4. Pupil-Linked Arousal and Neural Oscillations:

While future research needs to be conducted exploring each neuromodulatory system's direct role in neural oscillations, looking at another non-invasive metric, such as the pupil, could provide insight into the interaction between neuromodulation and cognition. Previous work has showcased the ability for the pupil to be used as a non-invasive readout of the central arousal state (pupil-linked arousal), with possible involvement of the noradrenergic and cholinergic systems [29,30,32,105,156]. In both humans and rodents, the pupil has been found to have a clear role in representing phasic arousal levels in perceptual decision-making tasks [34,157–161]. While studies have not yet explored the direct relationship between pupil and cross-frequency coupling, many have started to use pupil/oculomotor dynamics as an additional readout of brain state. For example, a study showcased how CFC between the phase-locked amplitudes of the gamma band and delta band corresponded to success in a visual discrimination task [162]. However, they failed to find that eye movements have any significant effect on the cross-frequency coupling in the EEG. An additional study explored the pupil dynamics in LFP recordings in the lateral hypothalamus, revealing a clear correlation between pupil dilation events and delta power, opening the door to new questions around pupillary changes and brain state transitions [163]. These studies raised new questions about the role of pupil-linked arousal in mediating cognitive states, with pupil dynamics potentially serving as an indicator of transition between neural oscillatory patterns resulting from neuromodulation.

5. Future Neuromodulation Technology

The functional outcomes of understanding the unique role each neuromodulatory system plays in mediating cross-frequency coupling to facilitate certain brain functions have immense potential. Already, researchers have been able to develop neural interfaces to control and modulate a variety of neurological pathways [164]. More modern techniques that pay attention to multiple modalities of data are now employing LFP or EEG recordings in conjunction with neural stimulation technologies, including vagus nerve stimulation (VNS), transcranial direct current stimulation, and ultrasound stimulation [165–171]. Techniques such as focused ultrasound stimulation have the potential of delivering localized brain stimulation, allowing for non-invasive selective activation of smaller brain regions such as the LC, BF, or VTA [169,172]. Likewise, more invasive techniques such as CNS microstimulation with carefully designed patterns have enabled preferential activation of axons and somas, permitting a targeted approach to the activation of local neuronal circuits [173–176]. Additional invasive approaches are also currently in development, with clinical trials of optogenetics in humans being explored as a method to treat pain through selective neuronal activation, as well as through the activation of retinal ganglion cells [177]. While invasive methods will always provide the greatest specificity in neuronal activation, other non-invasive stimulation approaches are also proving to be effective. Both tDCS and TMS have been seen as appropriate treatments for depression and addiction, modulating prefrontal regions linked to cognitive symptoms [178,179]. Recently, studies have been utilizing current steering in conjunction with tDCS, enabling selective activation of deeper brain structures without interfering with superficial cortical regions [180,181].

While these methods are promising, VNS is proving to be one of the most effective approaches to non-invasive neuromodulation. The vagus nerve projects to a variety of brain regions, including neuromodulatory centers such as the LC, and VNS has been shown to be involved in phase-amplitude coupling [18,182]. These questions open the

door to potentially developing the therapeutic paradigm in which VNS is conducted, using real-time neural oscillatory feedback as a form of optimization [18]. Additionally, other noninvasive stimulation methods, including transcutaneous vagus nerve stimulation, were shown to mimic VNS by modulating alpha EEG activity, suggesting that EEG can provide higher real-time feedback regarding arousal and cognitive changes [19,183]. Future adaptations of stimulation devices may need to incorporate time-variant phase–amplitude coupling, which could inform and decipher the role CFC plays in mediating or reflecting nervous system function [21]. While these enhancements may seem quite positive, one study showcased how theta–gamma cross frequency coupling induced via transcranial altering current stimulation actually worsened the ability of humans to employ cognitive control in goal-directed behavior [184–186]. It is essential for future devices, which aim to augment brain functions through the control of cognitive states, to optimize stimulation patterns based on real-time neural oscillatory dynamics to enable more informed treatments and feedback.

Future work reliably assessing real time cross-frequency coupling relationships in EEG and LFP signals could provide insight into how stimulation techniques can properly modulate cognitive function and behavioral outcomes. Experiments looking at key EEG features such as phase, amplitude, power, and frequency should be conducted in order to further explain how neural oscillations change in response to different cognitive processes. Selective neuromodulation of the noradrenergic, cholinergic, and dopaminergic system may provide insight into how different CFC phenomena occur, guiding our understanding into how complex synchronous neural networks communicate and formulate complex circuits to direct cognitive functions [187]. Closed-loop stimulation of these centers may provide a solution to uncovering the causality of CFC throughout the brain [9]. Neurological disease research for Parkinson’s, Alzheimer’s disease, schizophrenia, ADHD, and epilepsy may benefit greatly from this research, as neuromodulation could be used to change neural oscillatory patterns and network synchrony, potentially restoring or reversing erratic neural behavior caused by adverse neural synchronization [188].

6. Conclusions

The role of the neuromodulatory systems in influencing neural oscillations is an understudied topic. Decades of research have helped bring to light the relationship between neural oscillations and higher cognitive functions, but the causality behind these signals remains poorly understood. By reviewing the literature of each of the three major neuromodulatory systems (noradrenergic, cholinergic, and dopaminergic) and their representative cortical projections, effects on neural oscillations, and involvement in cross-frequency coupling, the complex nature of these neural computations can hopefully be further explained. To enhance our understanding of this relationship, it is essential to look towards specific oscillatory features such as cross-frequency coupling to understand the intricacies within large-scale neural synchronization. Studies emphasizing the importance of the activation and inhibition of these neuromodulatory centers in conjunction with frequency domain analysis could prove critical to understanding the causality of different behaviors and neurological disorders. These results could help inform the next generation of closed-loop neural stimulation devices with effective real-time outcomes, prioritizing the role of neuromodulation in controlling large-scale neural oscillations to restore or enhance brain functions.

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References

1. Buzsáki, G.; Draguhn, A. Neuronal Oscillations in Cortical Networks. *Science* **2004**, *304*, 1926–1929. <https://doi.org/10.1126/science.1099745>.
2. Fuster, J. *Cortex and mind: unifying cognition*; Oxford University Press: 2005.
3. Delis, I.; Ince, R.A.A.; Sajda, P.; Wang, Q. Neural encoding of active multi-sensing enhances perceptual decision-making via a synergistic cross-modal interaction. *J. Neurosci.* **2022**, *42*, 2344–2355. <https://doi.org/10.1523/jneurosci.0861-21.2022>.
4. Buzsáki, G.; Anastassiou, C.A.; Koch, C. The origin of extracellular fields and currents—EEG, ECoG, LFP and spikes. *Nat. Rev. Neurosci.* **2012**, *13*, 407–420. <https://doi.org/10.1038/nrn3241>.
5. Delis, I.; Dmochowski, J.P.; Sajda, P.; Wang, Q. Correlation of neural activity with behavioral kinematics reveals distinct sensory encoding and evidence accumulation processes during active tactile sensing. *NeuroImage* **2018**, *175*, 12–21. <https://doi.org/10.1016/j.neuroimage.2018.03.035>.
6. Michel, C.; Lehmann, D.; Henggeler, B.; Brandeis, D. Localization of the sources of EEG delta, theta, alpha and beta frequency bands using the FFT dipole approximation. *Electroencephalogr. Clin. Neurophysiol.* **1992**, *82*, 38–44. [https://doi.org/10.1016/0013-4694\(92\)90180-p](https://doi.org/10.1016/0013-4694(92)90180-p).
7. Friston, K.J. LFP and oscillations—what do they tell us? *Curr. Opin. Neurobiol.* **2015**, *31*, 1–6. <https://doi.org/10.1016/j.conb.2014.05.004>.
8. Hyafil, A.; Giraud, A.-L.; Fontolan, L.; Gutkin, B. Neural Cross-Frequency Coupling: Connecting Architectures, Mechanisms, and Functions. *Trends Neurosci.* **2015**, *38*, 725–740. <https://doi.org/10.1016/j.tins.2015.09.001>.
9. Salimpour, Y.; Anderson, W.S. Cross-Frequency Coupling Based Neuromodulation for Treating Neurological Disorders. *Front. Neurosci.* **2019**, *13*, 125. <https://doi.org/10.3389/fnins.2019.00125>.
10. Mormann, F.; Fell, J.; Axmacher, N.; Weber, B.; Lehnertz, K.; Elger, C.E.; Fernández, G. Phase/amplitude reset and theta–gamma interaction in the human medial temporal lobe during a continuous word recognition memory task. *Hippocampus* **2005**, *15*, 890–900. <https://doi.org/10.1002/hipo.20117>.
11. Yao, Y.; Wu, M.; Wang, L.; Lin, L.; Xu, J. Phase Coupled Firing of Prefrontal Parvalbumin Interneuron With High Frequency Oscillations. *Front. Cell. Neurosci.* **2020**, *14*, 610741. <https://doi.org/10.3389/fncel.2020.610741>.
12. Womelsdorf, T.; Schoffelen, J.M.; Oostenveld, R.; Singer, W.; Desimone, R.; Engel, A.K.; Fries, P. Modulation of neuronal interactions through neuronal synchronization. *Science* **2007**, *316*, 1609–1612. <https://doi.org/10.1126/science.1139597>.
13. Cavanagh, J.F.; Cohen, M.X.; Allen, J.J. Prelude to and resolution of an error: EEG phase synchrony reveals cognitive control dynamics during action monitoring. *J. Neurosci.* **2009**, *29*, 98–105. <https://doi.org/10.1523/JNEUROSCI.4137-08.2009>.
14. Fell, J.; Ludowig, E.; Rosburg, T.; Axmacher, N.; Elger, C.E. Phase-locking within human mediotemporal lobe predicts memory formation. *Neuroimage* **2008**, *43*, 410–419. <https://doi.org/10.1016/j.neuroimage.2008.07.021>.
15. Fell, J.; Axmacher, N. The role of phase synchronization in memory processes. *Nat. Rev. Neurosci.* **2011**, *12*, 105–118. <https://doi.org/10.1038/nrn2979>.
16. Başar, E. Brain oscillations in neuropsychiatric disease. *Dialogues Clin. Neurosci.* **2013**, *15*, 291–300. <https://doi.org/10.31887/DCNS.2013.15.3/ebasar>.
17. Salimpour, Y.; Mills, K.A.; Hwang, B.Y.; Anderson, W.S. Phase- targeted stimulation modulates phase-amplitude coupling in the motor cortex of the human brain. *Brain Stimul.* **2022**, *15*, 152–163. <https://doi.org/10.1016/j.brs.2021.11.019>.
18. Warsi, N.M.; Yan, H.; Wong, S.M.; Yau, I.; Breitbart, S.; Go, C.; Gorodetsky, C.; Fasano, A.; Kalia, S.K.; Rutka, J.T.; et al. Vagus Nerve Stimulation Modulates Phase-Amplitude Coupling in Thalamic Local Field Potentials. *Neuromodulation* **2022**, *in press*. <https://doi.org/10.1016/j.neurom.2022.05.001>.
19. Rodenkirch, C.; Carmel, J.B.; Wang, Q. Rapid Effects of Vagus Nerve Stimulation on Sensory Processing Through Activation of Neuromodulatory Systems. *Front. Neurosci.* **2022**, *16*, 922424. <https://doi.org/10.3389/fnins.2022.922424>.
20. Hulse, D.R.; Riley, J.R.; Loerwald, K.W.; Rennaker, R.L.; Kilgard, M.P.; Hays, S.A. Parametric characterization of neural activity in the locus coeruleus in response to vagus nerve stimulation. *Exp. Neurol.* **2017**, *289*, 21–30. <https://doi.org/10.1016/j.expneurol.2016.12.005>.

21. Chen, L.L.; Madhavan, R.; Rapoport, B.I.; Anderson, W.S. A method for real-time cortical oscillation detection and phase-locked stimulation. In Proceedings of the 2011 Annual International Conference of the IEEE Engineering in Medicine and Biology Society, Boston, MA, USA, 30 August–3 September 2011; Volume 2011, pp. 3087–3090. <https://doi.org/10.1109/IEMBS.2011.6090843>.
22. Descarries, L.; Watkins, K.C.; Lapierre, Y. Noradrenergic axon terminals in the cerebral cortex of rat. III. Topometric ultrastructural analysis. *Brain Res.* **1977**, *133*, 197–222. [https://doi.org/10.1016/0006-8993\(77\)90759-4](https://doi.org/10.1016/0006-8993(77)90759-4).
23. Li, X.; Yu, B.; Sun, Q.; Zhang, Y.; Ren, M.; Zhang, X.; Li, A.; Yuan, J.; Madisen, L.; Luo, Q.; et al. Generation of a whole-brain atlas for the cholinergic system and mesoscopic projectome analysis of basal forebrain cholinergic neurons. *Proc. Natl. Acad. Sci. USA* **2018**, *115*, 415–420. <https://doi.org/10.1073/pnas.1703601115>.
24. Björklund, A.; Dunnett, S.B. Dopamine neuron systems in the brain: An update. *Trends Neurosci.* **2007**, *30*, 194–202. <https://doi.org/10.1016/j.tins.2007.03.006>.
25. Rodenkirch, C.; Liu, Y.; Schriver, B.J.; Wang, Q. Locus coeruleus activation enhances thalamic feature selectivity via norepinephrine regulation of intrathalamic circuit dynamics. *Nat. Neurosci.* **2019**, *22*, 120–133. <https://doi.org/10.1038/s41593-018-0283-1>.
26. Slater, C.; Liu, Y.; Weiss, E.; Yu, K.; Wang, Q. The Neuromodulatory Role of the Noradrenergic and Cholinergic Systems and Their Interplay in Cognitive Functions: A Focused Review. *Brain Sci.* **2022**, *12*, 890. <https://doi.org/10.3390/brainsci12070890>.
27. Foote, S.L.; Bloom, F.E. Activity of Norepinephrine-Containing Locus Coeruleus Neurons in the Unanesthetized Squirrel Monkey. In *Catecholamines: Basic and Clinical Frontiers*; Usdin, E., Kopin, I.J., Barchas, J., Eds.; Pergamon, Turkey, 1979; pp. 625–627.
28. Totah, N.K.; Logothetis, N.K.; Eschenko, O. Synchronous spiking associated with prefrontal high γ oscillations evokes a 5-Hz rhythmic modulation of spiking in locus coeruleus. *J. Neurophysiol.* **2021**, *125*, 1191–1201. <https://doi.org/10.1152/jn.00677.2020>.
29. Szabadi, E. Functional Organization of the Sympathetic Pathways Controlling the Pupil: Light-Inhibited and Light-Stimulated Pathways. *Front. Neurol.* **2018**, *9*, 1069. <https://doi.org/10.3389/fneur.2018.01069>.
30. Joshi, S.; Li, Y.; Kalwani, R.M.; Gold, J.I. Relationships between Pupil Diameter and Neuronal Activity in the Locus Coeruleus, Colliculi, and Cingulate Cortex. *Neuron* **2016**, *89*, 221–234. <https://doi.org/10.1016/j.neuron.2015.11.028>.
31. Reimer, J.; Froudarakis, E.; Cadwell, C.R.; Yatsenko, D.; Denfield, G.H.; Tolias, A.S. Pupil fluctuations track fast switching of cortical states during quiet wakefulness. *Neuron* **2014**, *84*, 355–362. <https://doi.org/10.1016/j.neuron.2014.09.033>.
32. Reimer, J.; McGinley, M.J.; Liu, Y.; Rodenkirch, C.; Wang, Q.; McCormick, D.A.; Tolias, A.S. Pupil fluctuations track rapid changes in adrenergic and cholinergic activity in cortex. *Nat. Commun.* **2016**, *7*, 13289. <https://doi.org/10.1038/ncomms13289>.
33. McGinley, M.J.; David, S.V.; McCormick, D.A. Cortical membrane potential signature of optimal states for sensory signal detection. *Neuron* **2015**, *87*, 179–192. <https://doi.org/10.1016/j.neuron.2015.05.038>.
34. Liu, Y.; Narasimhan, S.; Schriver, B.J.; Wang, Q. Perceptual Behavior Depends Differently on Pupil-Linked Arousal and Heartbeat Dynamics-Linked Arousal in Rats Performing Tactile Discrimination Tasks. *Front. Syst. Neurosci.* **2021**, *14*, 614248. <https://doi.org/10.3389/fnsys.2020.614248>.
35. Saby, J.N.; Marshall, P.J. The Utility of EEG Band Power Analysis in the Study of Infancy and Early Childhood. *Dev. Neuropsychol.* **2012**, *37*, 253–273. <https://doi.org/10.1080/87565641.2011.614663>.
36. Stefanics, G.; Hangya, B.; Hernádi, I.; Winkler, I.; Lakatos, P.; Ulbert, I. Phase Entrainment of Human Delta Oscillations Can Mediate the Effects of Expectation on Reaction Speed. *J. Neurosci.* **2010**, *30*, 13578–13585. <https://doi.org/10.1523/JNEUROSCI.0703-10.2010>.
37. Amzica, F.; Steriade, M. Electrophysiological correlates of sleep delta waves. *Electroencephalogr. Clin. Neurophysiol.* **1998**, *107*, 69–83. [https://doi.org/10.1016/s0013-4694\(98\)00051-0](https://doi.org/10.1016/s0013-4694(98)00051-0).
38. Senoussi, M.; Verbeke, P.; Desender, K.; De Loof, E.; Talsma, D.; Verguts, T. Theta oscillations shift towards optimal frequency for cognitive control. *Nat. Hum. Behav.* **2022**, *6*, 1000–1013. <https://doi.org/10.1038/s41562-022-01335-5>.
39. Herweg, N.A.; Solomon, E.A.; Kahana, M.J. Theta Oscillations in Human Memory. *Trends Cogn. Sci.* **2020**, *24*, 208–227. <https://doi.org/10.1016/j.tics.2019.12.006>.
40. Lisman, J.E.; Jensen, O. The Theta-Gamma Neural Code. *Neuron* **2013**, *77*, 1002–1016. <https://doi.org/10.1016/j.neuron.2013.03.007>.
41. Lisman, J.; Buzsáki, G. A neural coding scheme formed by the combined function of gamma and theta oscillations. *Schizophr. Bull.* **2008**, *34*, 974–980. <https://doi.org/10.1093/schbul/sbn060>.
42. Kolev, V.; Yordanova, J. Analysis of phase-locking is informative for studying event-related EEG activity. *Biol. Cybern.* **1997**, *76*, 229–235. <https://doi.org/10.1007/s004220050335>.
43. Başar, E.; Schürmann, M.; Başar-Eroglu, C.; Karakaş, S. Alpha oscillations in brain functioning: An integrative theory. *Int. J. Psychophysiol.* **1997**, *26*, 5–29. [https://doi.org/10.1016/S0167-8760\(97\)00753-8](https://doi.org/10.1016/S0167-8760(97)00753-8).
44. Klimesch, W. Alpha-band oscillations, attention, and controlled access to stored information. *Trends Cogn. Sci.* **2012**, *16*, 606–617. <https://doi.org/10.1016/j.tics.2012.10.007>.
45. Cooper, N.R.; Croft, R.J.; Dominey, S.J.J.; Burgess, A.P.; Gruzelić, J.H. Paradox lost? Exploring the role of alpha oscillations during externally vs. internally directed attention and the implications for idling and inhibition hypotheses. *Int. J. Psychophysiol.* **2003**, *47*, 65–74. [https://doi.org/10.1016/S0167-8760\(02\)00107-1](https://doi.org/10.1016/S0167-8760(02)00107-1).
46. Busch, N.A.; Dubois, J.; VanRullen, R. The phase of ongoing EEG oscillations predicts visual perception. *J. Neurosci.* **2009**, *29*, 7869–7876. <https://doi.org/10.1523/JNEUROSCI.0113-09.2009>.

47. Foxe, J.J.; Snyder, A.C. The Role of Alpha-Band Brain Oscillations as a Sensory Suppression Mechanism during Selective Attention. *Front. Psychol.* **2011**, *2*, 154. <https://doi.org/10.3389/fpsyg.2011.00154>.
48. Pittman-Polletta, B.R.; Quach, A.; Mohammed, A.I.; Romano, M.; Kondabolu, K.; Kopell, N.J.; Han, X.; McCarthy, M.M. Striatal cholinergic receptor activation causes a rapid, selective and state-dependent rise in cortico-striatal β activity. *Eur. J. Neurosci.* **2018**, *48*, 2857–2868. <https://doi.org/10.1111/ejn.13906>.
49. Jenkinson, N.; Brown, P. New insights into the relationship between dopamine, beta oscillations and motor function. *Trends Neurosci.* **2011**, *34*, 611–618. <https://doi.org/10.1016/j.tins.2011.09.003>.
50. Schmidt, R.; Herrojo Ruiz, M.; Kilavik, B.E.; Lundqvist, M.; Starr, P.A.; Aron, A.R. Beta Oscillations in Working Memory, Executive Control of Movement and Thought, and Sensorimotor Function. *J. Neurosci.* **2019**, *39*, 8231–8238. <https://doi.org/10.1523/JNEUROSCI.1163-19.2019>.
51. Yang, Y.; Gritton, H.; Sarter, M.; Aton, S.J.; Booth, V.; Zochowski, M. Theta-gamma coupling emerges from spatially heterogeneous cholinergic neuromodulation. *PLoS Comput. Biol.* **2021**, *17*, e1009235. <https://doi.org/10.1371/journal.pcbi.1009235>.
52. Doesburg, S.; Vinette, S.; Cheung, M.; Pang, E. Theta-Modulated Gamma-Band Synchronization among Activated Regions During a Verb Generation Task. *Front. Psychol.* **2012**, *3*, 195. <https://doi.org/10.3389/fpsyg.2012.00195>.
53. Csicsvari, J.; Jamieson, B.; Wise, K.D.; Buzsáki, G. Mechanisms of Gamma Oscillations in the Hippocampus of the Behaving Rat. *Neuron* **2003**, *37*, 311–322. [https://doi.org/10.1016/S0896-6273\(02\)01169-8](https://doi.org/10.1016/S0896-6273(02)01169-8).
54. Crowley, K. Differentiating Pathologic Delta From Healthy Physiologic Delta in Patients With Alzheimer Disease. *Sleep* **2005**, *28*, 865–870. <https://doi.org/10.1093/sleep/28.7.865>.
55. Hiatt, J.F.; Floyd, T.C.; Katz, P.H.; Feinberg, I. Further Evidence of Abnormal Non-Rapid-Eye-Movement Sleep in Schizophrenia. *Arch. Gen. Psychiatry* **1985**, *42*, 797–802. <https://doi.org/10.1001/archpsyc.1985.01790310059007>.
56. Linkenkaer-Hansen, K.; Monto, S.; Rytälä, H.; Suominen, K.; Isometsä, E.; Kähkönen, S. Breakdown of Long-Range Temporal Correlations in Theta Oscillations in Patients with Major Depressive Disorder. *J. Neurosci.* **2005**, *25*, 10131–10137. <https://doi.org/10.1523/JNEUROSCI.3244-05.2005>.
57. Guo, J. Abnormal modulation of theta oscillations in children with attention-deficit/hyperactivity disorder. *NeuroImage Clin.* **2020**, *27*, 102314. <https://doi.org/10.1016/j.nicl.2020.102314>.
58. Osipova, D. Altered generation of spontaneous oscillations in Alzheimer's disease. *NeuroImage* **2005**, *27*, 835–841. <https://doi.org/10.1016/j.neuroimage.2005.05.011>.
59. Deiber, M.-P. Linking alpha oscillations, attention and inhibitory control in adult ADHD with EEG neurofeedback. *NeuroImage Clin.* **2020**, *25*, 102145. <https://doi.org/10.1016/j.nicl.2019.102145>.
60. Little, S. The functional role of beta oscillations in Parkinson's disease. *Park. Relat. Disord.* **2014**, *20*, S44–S48. [https://doi.org/10.1016/S1353-8020\(13\)70013-0](https://doi.org/10.1016/S1353-8020(13)70013-0).
61. Swann, N.C.; de Hemptinne, C.; Miocinovic, S.; Qasim, S.; Wang, S.S.; Ziman, N.; Ostrem, J.L.; San Luciano, M.; Galifianakis, N.B.; Starr, P.A. Gamma Oscillations in the Hyperkinetic State Detected with Chronic Human Brain Recordings in Parkinson's Disease. *J. Neurosci.* **2016**, *36*, 6445–6458. <https://doi.org/10.1523/JNEUROSCI.1128-16.2016>.
62. Mably, A.J. Gamma oscillations in cognitive disorders. *Curr. Opin. Neurobiol.* **2018**, *52*, 182–187. <https://doi.org/10.1016/j.conb.2018.07.009>.
63. Palva, S.; Palva, J.M. New vistas for α -frequency band oscillations. *Trends Neurosci.* **2007**, *30*, 150–158. <https://doi.org/10.1016/j.tins.2007.02.001>.
64. Yakubov, B.; Das, S.; Zomorodi, R.; Blumberger, D.M.; Enticott, P.G.; Kirkovski, M.; Rajji, T.K.; Desarkar, P. Cross-frequency coupling in psychiatric disorders: A systematic review. *Neurosci. Biobehav. Rev.* **2022**, *138*, 104690. <https://doi.org/10.1016/j.neubiorev.2022.104690>.
65. Tort, A.B.; Komorowski, R.W.; Manns, J.R.; Kopell, N.J.; Eichenbaum, H. Theta-gamma coupling increases during the learning of item-context associations. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 20942–20947. <https://doi.org/10.1073/pnas.0911331106>.
66. Nadalin, J.K.; Martinet, L.E.; Blackwood, E.B.; Lo, M.C.; Widge, A.S.; Cash, S.S.; Eden, U.T.; Kramer, M.A. A statistical framework to assess cross-frequency coupling while accounting for confounding analysis effects. *Elife* **2019**, *8*, e44287. <https://doi.org/10.7554/eLife.44287>.
67. Ceni, A.; Olmi, S.; Torcini, A.; Angulo-Garcia, D. Cross frequency coupling in next generation inhibitory neural mass models. *Chaos* **2020**, *30*, 053121. <https://doi.org/10.1063/1.5125216>.
68. Buzsáki, G.; Wang, X.-J. Mechanisms of gamma oscillations. *Annu. Rev. Neurosci.* **2012**, *35*, 203–225. <https://doi.org/10.1146/annurev-neuro-062111-150444>.
69. Jirsa, V.; Müller, V. Cross-frequency coupling in real and virtual brain networks. *Front. Comput. Neurosci.* **2013**, *7*, 78. <https://doi.org/10.3389/fncom.2013.00078>.
70. Cohen, M.X. Assessing transient cross-frequency coupling in EEG data. *J. Neurosci. Methods* **2008**, *168*, 494–499. <https://doi.org/10.1016/j.jneumeth.2007.10.012>.
71. Osipova, D.; Hermes, D.; Jensen, O. Gamma power is phase-locked to posterior alpha activity. *PLoS ONE* **2008**, *3*, e3990. <https://doi.org/10.1371/journal.pone.0003990>.
72. Andino-Pavlovsky, V.; Souza, A.C.; Scheffer-Teixeira, R.; Tort, A.B.L.; Etchenique, R.; Ribeiro, S. Dopamine Modulates Delta-Gamma Phase-Amplitude Coupling in the Prefrontal Cortex of Behaving Rats. *Front. Neural Circuits* **2017**, *11*, 29. <https://doi.org/10.3389/fncir.2017.00029>.

73. Cole, S.R.; Voytek, B. Brain Oscillations and the Importance of Waveform Shape. *Trends Cogn. Sci.* **2017**, *21*, 137–149. <https://doi.org/10.1016/j.tics.2016.12.008>.
74. Canolty, R.T.; Knight, R.T. The functional role of cross-frequency coupling. *Trends Cogn. Sci.* **2010**, *14*, 506–515. <https://doi.org/10.1016/j.tics.2010.09.001>.
75. Bandarabadi, M.; Boyce, R.; Gutierrez Herrera, C.; Bassetti, C.L.; Williams, S.; Schindler, K.; Adamantidis, A. Dynamic modulation of theta–gamma coupling during rapid eye movement sleep. *Sleep* **2019**, *42*, zsz182. <https://doi.org/10.1093/sleep/zsz182>.
76. Butler, J.L.; Paulsen, O. Hippocampal network oscillations—recent insights from in vitro experiments. *Curr. Opin. Neurobiol.* **2015**, *31*, 40–44. <https://doi.org/10.1016/j.conb.2014.07.025>.
77. Tamura, M.; Spellman, T.J.; Rosen, A.M.; Gogos, J.A.; Gordon, J.A. Hippocampal-prefrontal theta-gamma coupling during performance of a spatial working memory task. *Nat. Commun.* **2017**, *8*, 2182. <https://doi.org/10.1038/s41467-017-02108-9>.
78. Cohen, M.X.; Elger, C.E.; Fell, J. Oscillatory activity and phase-amplitude coupling in the human medial frontal cortex during decision making. *J. Cogn. Neurosci.* **2009**, *21*, 390–402. <https://doi.org/10.1162/jocn.2008.21020>.
79. Watrous, A.J.; Deuker, L.; Fell, J.; Axmacher, N. Phase-amplitude coupling supports phase coding in human ECoG. *Elife* **2015**, *4*, e07886. <https://doi.org/10.7554/eLife.07886>.
80. Seymour, R.A.; Rippon, G.; Kessler, K. The Detection of Phase Amplitude Coupling during Sensory Processing. *Front. Neurosci.* **2017**, *11*, 487. <https://doi.org/10.3389/fnins.2017.00487>.
81. Chacko, R.V.; Kim, B.; Jung, S.W.; Daitch, A.L.; Roland, J.L.; Metcalfe, N.V.; Corbetta, M.; Shulman, G.L.; Leuthardt, E.C. Distinct phase-amplitude couplings distinguish cognitive processes in human attention. *NeuroImage* **2018**, *175*, 111–121. <https://doi.org/10.1016/j.neuroimage.2018.03.003>.
82. Buschman, T.J.; Denovellis, E.L.; Diogo, C.; Bullock, D.; Miller, E.K. Synchronous Oscillatory Neural Ensembles for Rules in the Prefrontal Cortex. *Neuron* **2012**, *76*, 838–846. <https://doi.org/10.1016/j.neuron.2012.09.029>.
83. Szczepanski, S.M.; Crone, N.E.; Kuperman, R.A.; Augustine, K.I.; Parvizi, J.; Knight, R.T. Dynamic Changes in Phase-Amplitude Coupling Facilitate Spatial Attention Control in Fronto-Parietal Cortex. *PLoS Biol.* **2014**, *12*, e1001936. <https://doi.org/10.1371/journal.pbio.1001936>.
84. Rihs, T.A.; Michel, C.M.; Thut, G. Mechanisms of selective inhibition in visual spatial attention are indexed by α -band EEG synchronization. *Eur. J. Neurosci.* **2007**, *25*, 603–610. <https://doi.org/10.1111/j.1460-9568.2007.05278.x>.
85. Chen, S.; Tan, Z.; Xia, W.; Gomes, C.A.; Zhang, X.; Zhou, W.; Liang, S.; Axmacher, N.; Wang, L. Theta oscillations synchronize human medial prefrontal cortex and amygdala during fear learning. *Sci. Adv.* **2021**, *7*, eabf4198. <https://doi.org/10.1126/sciadv.abf4198>.
86. Klimesch, W. Memory processes, brain oscillations and EEG synchronization. *Int. J. Psychophysiol.* **1996**, *24*, 61–100. [https://doi.org/10.1016/S0167-8760\(96\)00057-8](https://doi.org/10.1016/S0167-8760(96)00057-8).
87. Siegel, M.; Warden, M.R.; Miller, E.K. Phase-dependent neuronal coding of objects in short-term memory. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 21341–21346. <https://doi.org/10.1073/pnas.0908193106>.
88. De Hemptinne, C.; Ryapolova-Webb, E.S.; Air, E.L.; Garcia, P.A.; Miller, K.J.; Ojemann, J.G.; Ostrem, J.L.; Galifianakis, N.B.; Starr, P.A. Exaggerated phase-amplitude coupling in the primary motor cortex in Parkinson disease. *Proc. Natl. Acad. Sci. USA* **2013**, *110*, 4780–4785. <https://doi.org/10.1073/pnas.1214546110>.
89. Goutagny, R.; Gu, N.; Cavanagh, C.; Jackson, J.; Chabot, J.-G.; Quirion, R.; Krantic, S.; Williams, S. Alterations in hippocampal network oscillations and theta-gamma coupling arise before A β overproduction in a mouse model of Alzheimer’s disease. *Eur. J. Neurosci.* **2013**, *37*, 1896–1902. <https://doi.org/10.1111/ejn.12233>.
90. Mirzayi, P.; Shobeiri, P.; Kalantari, A.; Perry, G.; Rezaei, N. Optogenetics: Implications for Alzheimer’s disease research and therapy. *Mol. Brain* **2022**, *15*, 20. <https://doi.org/10.1186/s13041-022-00905-y>.
91. Kim, J.W.; Lee, J.; Kim, B.-N.; Kang, T.; Min, K.J.; Han, D.H.; Lee, Y.S. Theta-phase gamma-amplitude coupling as a neurophysiological marker of attention deficit/hyperactivity disorder in children. *Neurosci. Lett.* **2015**, *603*, 25–30. <https://doi.org/10.1016/j.neulet.2015.07.006>.
92. Moran, L.V.; Hong, L.E. High vs. low frequency neural oscillations in schizophrenia. *Schizophr. Bull.* **2011**, *37*, 659–663. <https://doi.org/10.1093/schbul/sbr056>.
93. Kirihaara, K.; Rissling, A.J.; Swerdlow, N.R.; Braff, D.L.; Light, G.A. Hierarchical Organization of Gamma and Theta Oscillatory Dynamics in Schizophrenia. *Biol. Psychiatry* **2012**, *71*, 873–880. <https://doi.org/10.1016/j.biopsych.2012.01.016>.
94. Loizou, L.A. Projections of the nucleus locus coeruleus in the albino rat. *Brain Res.* **1969**, *15*, 563–566. [https://doi.org/10.1016/0006-8993\(69\)90185-1](https://doi.org/10.1016/0006-8993(69)90185-1).
95. Russell, G.V. The nucleus locus coeruleus (dorsolateralis tegmenti). *Tex. Rep. Biol. Med.* **1955**, *13*, 939–988.
96. Totah, N.K.; Neves, R.M.; Panzeri, S.; Logothetis, N.K.; Eschenko, O. The Locus Coeruleus Is a Complex and Differentiated Neuromodulatory System. *Neuron* **2018**, *99*, 1055–1068. <https://doi.org/10.1016/j.neuron.2018.05.056>.
97. Chandler, D.J.; Jensen, P.; McCall, J.G.; Pickering, A.E.; Schwarz, L.A.; Totah, N.K. Redefining Noradrenergic Neuromodulation of Behavior: Impacts of a Modular Locus Coeruleus Architecture. *J. Neurosci.* **2019**, *39*, 8239. <https://doi.org/10.1523/JNEUROSCI.1164-19.2019>.
98. Ross, J.A.; Van Bockstaele, E.J. The Locus Coeruleus- Norepinephrine System in Stress and Arousal: Unraveling Historical, Current, and Future Perspectives. *Front. Psychiatry* **2020**, *11*, 601519. <https://doi.org/10.3389/fpsy.2020.601519>.

99. Sara, S.J.; Bouret, S. Orienting and reorienting: The locus coeruleus mediates cognition through arousal. *Neuron* **2012**, *76*, 130–141. <https://doi.org/10.1016/j.neuron.2012.09.011>.
100. Bouret, S.; Sara, S.J. Locus coeruleus activation modulates firing rate and temporal organization of odour-induced single-cell responses in rat piriform cortex. *Eur. J. Neurosci.* **2002**, *16*, 2371–2382. <https://doi.org/10.1046/j.1460-9568.2002.02413.x>.
101. Grella, S.L.; Neil, J.M.; Edison, H.T.; Strong, V.D.; Odintsova, I.V.; Walling, S.G.; Martin, G.M.; Marrone, D.F.; Harley, C.W. Locus Coeruleus Phasic, But Not Tonic, Activation Initiates Global Remapping in a Familiar Environment. *J. Neurosci.* **2019**, *39*, 445–455. <https://doi.org/10.1523/JNEUROSCI.1956-18.2018>.
102. Aston-Jones, G.; Rajkowski, J.; Cohen, J. Role of locus coeruleus in attention and behavioral flexibility. *Biol. Psychiatry* **1999**, *46*, 1309–1320.
103. Hayat, H.; Regev, N.; Matosevich, N.; Sales, A.; Paredes-Rodriguez, E.; Krom, A.J.; Bergman, L.; Li, Y.; Lavigne, M.; Kremer, E.J.; et al. Locus coeruleus norepinephrine activity mediates sensory-evoked awakenings from sleep. *Sci. Adv.* **2020**, *6*, eaaz4232. <https://doi.org/10.1126/sciadv.aaz4232>.
104. Foote, S.L.; Aston-Jones, G.; Bloom, F.E. Impulse activity of locus coeruleus neurons in awake rats and monkeys is a function of sensory stimulation and arousal. *Proc. Natl. Acad. Sci. USA* **1980**, *77*, 3033–3037.
105. Liu, Y.; Rodenkirch, C.; Moskowitz, N.; Schriver, B.; Wang, Q. Dynamic Lateralization of Pupil Dilation Evoked by Locus Coeruleus Activation Results from Sympathetic, Not Parasympathetic, Contributions. *Cell Rep.* **2017**, *20*, 3099–3112. <https://doi.org/10.1016/j.celrep.2017.08.094>.
106. Broncel, A.; Bocian, R.; Kłos-Wojtczak, P.; Konopacki, J. Effects of locus coeruleus activation and inactivation on hippocampal formation theta rhythm in anesthetized rats. *Brain Res. Bull.* **2020**, *162*, 180–190. <https://doi.org/10.1016/j.brainresbull.2020.05.017>.
107. Lemon, N.; Aydin-Abidin, S.; Funke, K.; Manahan-Vaughan, D. Locus coeruleus activation facilitates memory encoding and induces hippocampal LTD that depends on beta-adrenergic receptor activation. *Cereb. Cortex* **2009**, *19*, 2827–2837. <https://doi.org/10.1093/cercor/bhp065>.
108. Geiller, T.; Sadeh, S.; Rolotti, S.V.; Blockus, H.; Vancura, B.; Negrean, A.; Murray, A.J.; Rozsa, B.; Polleux, F.; Clopath, C.; et al. Local circuit amplification of spatial selectivity in the hippocampus. *Nature* **2022**, *601*, 105–109. <https://doi.org/10.1038/s41586-021-04169-9>.
109. Natsume, K.; Kometani, K. Desynchronization of carbachol-induced theta-like activities by alpha-adrenergic agents in guinea pig hippocampal slices. *Neurosci. Res.* **1999**, *33*, 179–186. [https://doi.org/10.1016/s0168-0102\(99\)00007-3](https://doi.org/10.1016/s0168-0102(99)00007-3).
110. Hajós, M.; Hoffmann, W.E.; Robinson, D.D.; Yu, J.H.; Hajós-Korcsok, E. Norepinephrine but not serotonin reuptake inhibitors enhance theta and gamma activity of the septo-hippocampal system. *Neuropsychopharmacology* **2003**, *28*, 857–864. <https://doi.org/10.1038/sj.npp.1300116>.
111. Aston-Jones, G.; Cohen, J.D. Adaptive gain and the role of the locus coeruleus-norepinephrine system in optimal performance. *J. Comp. Neurol.* **2005**, *493*, 99–110. <https://doi.org/10.1002/cne.20723>.
112. Bouret, S.; Sara, S.J. Network reset: A simplified overarching theory of locus coeruleus noradrenaline function. *Trends Neurosci.* **2005**, *28*, 574–582. <https://doi.org/10.1016/j.tins.2005.09.002>.
113. McBurney-Lin, J.; Lu, J.; Zuo, Y.; Yang, H. Locus Coeruleus-Norepinephrine Modulation of Sensory Processing and Perception: A Focused Review. *Neurosci. Biobehav. Rev.* **2019**, *105*, 190–199. <https://doi.org/10.1016/j.neubiorev.2019.06.009>.
114. Dahl, M.J.; Mather, M.; Werkle-Bergner, M. Noradrenergic modulation of rhythmic neural activity shapes selective attention. *Trends Cogn. Sci.* **2022**, *26*, 38–52. <https://doi.org/10.1016/j.tics.2021.10.009>.
115. Durán, E.; Yang, M.; Neves, R.; Logothetis, N.K.; Eschenko, O. Modulation of Prefrontal Cortex Slow Oscillations by Phasic Activation of the Locus Coeruleus. *Neuroscience* **2021**, *453*, 268–279. <https://doi.org/10.1016/j.neuroscience.2020.11.028>.
116. Semba, K.; Fibiger, H.C. Organization of central cholinergic systems. *Prog. Brain Res.* **1989**, *79*, 37–63. [https://doi.org/10.1016/s0079-6123\(08\)62464-4](https://doi.org/10.1016/s0079-6123(08)62464-4).
117. Woolf, N.J.; Eckenstein, F.; Butcher, L.L. Cholinergic systems in the rat brain: I. projections to the limbic telencephalon. *Brain Res. Bull.* **1984**, *13*, 751–784. [https://doi.org/10.1016/0361-9230\(84\)90236-3](https://doi.org/10.1016/0361-9230(84)90236-3).
118. Kása, P.; Rakonczay, Z.; Gulya, K. The cholinergic system in Alzheimer's disease. *Prog. Neurobiol.* **1997**, *52*, 511–535. [https://doi.org/10.1016/s0301-0082\(97\)00028-2](https://doi.org/10.1016/s0301-0082(97)00028-2).
119. Bohnen, N.I.; Albin, R.L. The cholinergic system and Parkinson disease. *Behav. Brain Res.* **2011**, *221*, 564–573. <https://doi.org/10.1016/j.bbr.2009.12.048>.
120. Rye, D.B.; Wainer, B.H.; Mesulam, M.-M.; Mufson, E.J.; Saper, C.B. Cortical projections arising from the basal forebrain: A study of cholinergic and noncholinergic components employing combined retrograde tracing and immunohistochemical localization of choline acetyltransferase. *Neuroscience* **1984**, *13*, 627–643. [https://doi.org/10.1016/0306-4522\(84\)90083-6](https://doi.org/10.1016/0306-4522(84)90083-6).
121. Woolf, N.J. Cholinergic systems in mammalian brain and spinal cord. *Prog. Neurobiol.* **1991**, *37*, 475–524. [https://doi.org/10.1016/0301-0082\(91\)90006-m](https://doi.org/10.1016/0301-0082(91)90006-m).
122. Passetti, F.; Dalley, J.W.; O'Connell, M.T.; Everitt, B.J.; Robbins, T.W. Increased acetylcholine release in the rat medial prefrontal cortex during performance of a visual attentional task. *Eur. J. Neurosci.* **2000**, *12*, 3051–3058. <https://doi.org/10.1046/j.1460-9568.2000.00183.x>.
123. Sarter, M.; Parikh, V.; Howe, W.M. Phasic acetylcholine release and the volume transmission hypothesis: Time to move on. *Nat. Rev. Neurosci.* **2009**, *10*, 383–390. <https://doi.org/10.1038/nrn2635>.

124. Mattinson, C.E.; Burmeister, J.J.; Quintero, J.E.; Pomerleau, F.; Huettl, P.; Gerhardt, G.A. Tonic and phasic release of glutamate and acetylcholine neurotransmission in sub-regions of the rat prefrontal cortex using enzyme-based microelectrode arrays. *J. Neurosci. Methods* **2011**, *202*, 199–208. <https://doi.org/10.1016/j.jneumeth.2011.08.020>.
125. Teles-Grilo Ruivo, L.M.; Baker, K.L.; Conway, M.W.; Kinsley, P.J.; Gilmour, G.; Phillips, K.G.; Isaac, J.T.R.; Lowry, J.P.; Mellor, J.R. Coordinated Acetylcholine Release in Prefrontal Cortex and Hippocampus Is Associated with Arousal and Reward on Distinct Timescales. *Cell Rep.* **2017**, *18*, 905–917. <https://doi.org/10.1016/j.celrep.2016.12.085>.
126. Sarter, M.; Lustig, C.; Berry, A.S.; Gritton, H.; Howe, W.M.; Parikh, V. What do phasic cholinergic signals do? *Neurobiol. Learn. Mem.* **2016**, *130*, 135–141. <https://doi.org/10.1016/j.nlm.2016.02.008>.
127. Steriade, M. Acetylcholine systems and rhythmic activities during the waking–sleep cycle. *Prog. Brain Res.* **2004**, *145*, 179–196.
128. Goard, M.; Dan, Y. Basal forebrain activation enhances cortical coding of natural scenes. *Nat. Neurosci.* **2009**, *12*, 1444–1449.
129. Platt, B.; Riedel, G. The cholinergic system, EEG and sleep. *Behav. Brain Res.* **2011**, *221*, 499–504. <https://doi.org/10.1016/j.bbr.2011.01.017>.
130. Jones, B.E. Activity, modulation and role of basal forebrain cholinergic neurons innervating the cerebral cortex. *Prog. Brain Res.* **2004**, *145*, 157–169.
131. Cape, E.G.; Manns, I.D.; Alonso, A.; Beaudet, A.; Jones, B.E. Neurotensin-Induced Bursting of Cholinergic Basal Forebrain Neurons Promotes γ and θ Cortical Activity Together with Waking and Paradoxical Sleep. *J. Neurosci.* **2000**, *20*, 8452–8461. <https://doi.org/10.1523/JNEUROSCI.20-22-08452.2000>.
132. Vinogradova, O.S.; Brazhnik, E.S.; Kitchigina, V.F.; Stafekhina, V.S. Acetylcholine, theta-rhythm and activity of hippocampal neurons in the rabbit—IV. Sensory stimulation. *Neuroscience* **1993**, *53*, 993–1007. [https://doi.org/10.1016/0306-4522\(93\)90484-W](https://doi.org/10.1016/0306-4522(93)90484-W).
133. Hasselmo, M.E.; Hay, J.; Ilyn, M.; Gorchetchnikov, A. Neuromodulation, theta rhythm and rat spatial navigation. *Neural Networks* **2002**, *15*, 689–707. [https://doi.org/10.1016/S0893-6080\(02\)00057-6](https://doi.org/10.1016/S0893-6080(02)00057-6).
134. Hasselmo, M.E. The role of acetylcholine in learning and memory. *Curr. Opin. Neurobiol.* **2006**, *16*, 710–715. <https://doi.org/10.1016/j.conb.2006.09.002>.
135. Stojilkovic, M.; Kelley, C.; Nagy, D.; Leventhal, L.; Hajós, M. Selective activation of $\alpha 7$ nicotinic acetylcholine receptors augments hippocampal oscillations. *Neuropharmacology* **2016**, *110*, 102–108. <https://doi.org/10.1016/j.neuropharm.2016.07.010>.
136. Howe, W.M.; Gritton, H.J.; Lusk, N.A.; Roberts, E.A.; Hetrick, V.L.; Berke, J.D.; Sarter, M. Acetylcholine Release in Prefrontal Cortex Promotes Gamma Oscillations and Theta-Gamma Coupling during Cue Detection. *J. Neurosci.* **2017**, *37*, 3215–3230. <https://doi.org/10.1523/jneurosci.2737-16.2017>.
137. McNally, J.M.; Aguilar, D.D.; Katsuki, F.; Radzik, L.K.; Schiffino, F.L.; Uygun, D.S.; McKenna, J.T.; Strecker, R.E.; Deisseroth, K.; Spencer, K.M.; et al. Optogenetic manipulation of an ascending arousal system tunes cortical broadband gamma power and reveals functional deficits relevant to schizophrenia. *Mol. Psychiatry* **2021**, *26*, 3461–3475. <https://doi.org/10.1038/s41380-020-0840-3>.
138. Hwang, E.; Brown, R.E.; Kocsis, B.; Kim, T.; McKenna, J.T.; McNally, J.M.; Han, H.B.; Choi, J.H. Optogenetic stimulation of basal forebrain parvalbumin neurons modulates the cortical topography of auditory steady-state responses. *Brain Struct. Funct.* **2019**, *224*, 1505–1518. <https://doi.org/10.1007/s00429-019-01845-5>.
139. Neymotin, S.A.; Hilscher, M.M.; Moulin, T.C.; Skolnick, Y.; Lazarewicz, M.T.; Lytton, W.W. Ih tunes theta/gamma oscillations and cross-frequency coupling in an in silico CA3 model. *PLoS ONE* **2013**, *8*, e76285. <https://doi.org/10.1371/journal.pone.0076285>.
140. Rasmusson, D.D.; Clow, K.; Szerb, J.C. Modification of neocortical acetylcholine release and electroencephalogram desynchronization due to brainstem stimulation by drugs applied to the basal forebrain. *Neuroscience* **1994**, *60*, 665–677. [https://doi.org/10.1016/0306-4522\(94\)90495-2](https://doi.org/10.1016/0306-4522(94)90495-2).
141. Goldman-Rakic, P.S.; Leranth, C.; Williams, S.M.; Mons, N.; Geffard, M. Dopamine synaptic complex with pyramidal neurons in primate cerebral cortex. *Proc. Natl. Acad. Sci. USA* **1989**, *86*, 9015–9019. <https://doi.org/10.1073/pnas.86.22.9015>.
142. Ott, T.; Nieder, A. Dopamine and Cognitive Control in Prefrontal Cortex. *Trends Cogn. Sci.* **2019**, *23*, 213–234. <https://doi.org/10.1016/j.tics.2018.12.006>.
143. Williams, S.M.; Goldman-Rakic, P.S. Widespread origin of the primate mesofrontal dopamine system. *Cereb. Cortex* **1998**, *8*, 321–345. <https://doi.org/10.1093/cercor/8.4.321>.
144. Dreyer, J.K.; Herrik, K.F.; Berg, R.W.; Hounsgaard, J.D. Influence of Phasic and Tonic Dopamine Release on Receptor Activation. *J. Neurosci.* **2010**, *30*, 14273–14283. <https://doi.org/10.1523/JNEUROSCI.1894-10.2010>.
145. Puig, M.V.; Miller, E.K. The Role of Prefrontal Dopamine D1 Receptors in the Neural Mechanisms of Associative Learning. *Neuron* **2012**, *74*, 874–886. <https://doi.org/10.1016/j.neuron.2012.04.018>.
146. Sharott, A.; Magill, P.J.; Harnack, D.; Kupsch, A.; Meissner, W.; Brown, P. Dopamine depletion increases the power and coherence of beta-oscillations in the cerebral cortex and subthalamic nucleus of the awake rat. *Eur. J. Neurosci.* **2005**, *21*, 1413–1422. <https://doi.org/10.1111/j.1460-9568.2005.03973.x>.
147. Eckart, C.; Fuentemilla, L.; Bauch, E.M.; Bunzeck, N. Dopaminergic stimulation facilitates working memory and differentially affects prefrontal low theta oscillations. *NeuroImage* **2014**, *94*, 185–192. <https://doi.org/10.1016/j.neuroimage.2014.03.011>.
148. Benchenane, K.; Peyrache, A.; Khamassi, M.; Tierney, P.L.; Gioanni, Y.; Battaglia, F.P.; Wiener, S.I. Coherent Theta Oscillations and Reorganization of Spike Timing in the Hippocampal- Prefrontal Network upon Learning. *Neuron* **2010**, *66*, 921–936. <https://doi.org/10.1016/j.neuron.2010.05.013>.

149. Taylor, N.E.; Van Dort, C.J.; Kenny, J.D.; Pei, J.; Guidera, J.A.; Vlasov, K.Y.; Lee, J.T.; Boyden, E.S.; Brown, E.N.; Solt, K. Optogenetic activation of dopamine neurons in the ventral tegmental area induces reanimation from general anesthesia. *Proc. Natl. Acad. Sci. USA* **2016**, *113*, 12826–12831. <https://doi.org/10.1073/pnas.1614340113>.
150. Mallet, N.; Pogosyan, A.; Sharott, A.; Csicsvari, J.; Bolam, J.P.; Brown, P.; Magill, P.J. Disrupted dopamine transmission and the emergence of exaggerated beta oscillations in subthalamic nucleus and cerebral cortex. *J. Neurosci.* **2008**, *28*, 4795–4806. <https://doi.org/10.1523/JNEUROSCI.0123-08.2008>.
151. Iskhakova, L.; Rappel, P.; Deffains, M.; Fonar, G.; Marmor, O.; Paz, R.; Israel, Z.; Eitan, R.; Bergman, H. Modulation of dopamine tone induces frequency shifts in cortico-basal ganglia beta oscillations. *Nat. Commun.* **2021**, *12*, 7026. <https://doi.org/10.1038/s41467-021-27375-5>.
152. Degos, B.; Deniau, J.M.; Chavez, M.; Maurice, N. Chronic but not acute dopaminergic transmission interruption promotes a progressive increase in cortical beta frequency synchronization: Relationships to vigilance state and akinesia. *Cereb. Cortex* **2009**, *19*, 1616–1630. <https://doi.org/10.1093/cercor/bhn199>.
153. Ongini, E.; Caporali, M.G.; Massotti, M. Stimulation of dopamine D-1 receptors by SKF 38393 induces EEG desynchronization and behavioral arousal. *Life Sci.* **1985**, *37*, 2327–2333. [https://doi.org/10.1016/0024-3205\(85\)90025-6](https://doi.org/10.1016/0024-3205(85)90025-6).
154. Xu, X.; Zheng, C.; An, L.; Wang, R.; Zhang, T. Effects of Dopamine and Serotonin Systems on Modulating Neural Oscillations in Hippocampus-Prefrontal Cortex Pathway in Rats. *Brain Topogr.* **2016**, *29*, 539–551. <https://doi.org/10.1007/s10548-016-0485-3>.
155. Lohani, S.; Martig, A.K.; Deisseroth, K.; Witten, I.B.; Moghaddam, B. Dopamine Modulation of Prefrontal Cortex Activity Is Manifold and Operates at Multiple Temporal and Spatial Scales. *Cell Rep.* **2019**, *27*, 99–114.e116. <https://doi.org/10.1016/j.celrep.2019.03.012>.
156. Carter, M.E.; Yizhar, O.; Chikahisa, S.; Nguyen, H.; Adamantidis, A.; Nishino, S.; Deisseroth, K.; de Lecea, L. Tuning arousal with optogenetic modulation of locus coeruleus neurons. *Nat. Neurosci.* **2010**, *13*, 1526–1533. <https://doi.org/10.1038/nn.2682>.
157. Schriver, B.J.; Perkins, S.M.; Sajda, P.; Wang, Q. Interplay between components of pupil-linked phasic arousal and its role in driving behavioral choice in Go/No-Go perceptual decision-making. *Psychophysiology* **2020**, *57*, e13565. <https://doi.org/10.1111/psyp.13565>.
158. de Gee, J.W.; Tsetsos, K.; Schwabe, L.; Urai, A.E.; McCormick, D.; McGinley, M.J.; Donner, T.H. Pupil-linked phasic arousal predicts a reduction of choice bias across species and decision domains. *eLife* **2020**, *9*, e54014. <https://doi.org/10.7554/eLife.54014>.
159. Schriver, B.; Bagdasarov, S.; Wang, Q. Pupil-linked arousal modulates behavior in rats performing a whisker deflection direction discrimination task. *J. Neurophysiol.* **2018**, *120*, 1655–1670. <https://doi.org/10.1152/jn.00290.2018>.
160. Lapborisuth, P.; Koorathota, S.; Wang, Q.; Sajda, P. Integrating neural and ocular attention reorienting signals in virtual reality. *J. Neural Eng.* **2022**, *18*, 066052. <https://doi.org/10.1088/1741-2552/ac4593>.
161. Narasimhan, S.; Schriver, B.J.; Wang, Q. Adaptive decision making depends on pupil-linked arousal in rats performing tactile discrimination tasks. *bioRxiv* **2022**, <https://doi.org/10.1101/2022.07.20.500875>.
162. Händel, B.; Haarmeier, T. Cross-frequency coupling of brain oscillations indicates the success in visual motion discrimination. *NeuroImage* **2009**, *45*, 1040–1046. <https://doi.org/10.1016/j.neuroimage.2008.12.013>.
163. Takahashi, K.; Sobczak, F.; Pais-Roldán, P.; Yu, X. Characterizing pupil dynamics coupling to brain state fluctuation based on lateral hypothalamic activity. *bioRxiv* **2021**, <https://doi.org/10.1101/2021.09.22.461385>.
164. Rodenkirch, C.; Schriver, B.; Wang, Q. Brain-Machine Interfaces: Restoring and Establishing Communication Channels. In *Neural Engineering: From Advanced Biomaterials to 3D Fabrication Techniques*; Zhang, L., Kaplan, D., Eds.; Springer: Cham, Switzerland, 2016.
165. Park, M.; Hoang, G.M.; Nguyen, T.; Lee, E.; Jung, H.J.; Choe, Y.; Lee, M.H.; Hwang, J.Y.; Kim, J.G.; Kim, T. Effects of transcranial ultrasound stimulation pulsed at 40 Hz on Aβ plaques and brain rhythms in 5×FAD mice. *Transl. Neurodegener.* **2021**, *10*, 48. <https://doi.org/10.1186/s40035-021-00274-x>.
166. Thut, G.; Schyns, P.G.; Gross, J. Entrainment of perceptually relevant brain oscillations by non-invasive rhythmic stimulation of the human brain. *Front. Psychol.* **2011**, *2*, 170. <https://doi.org/10.3389/fpsyg.2011.00170>.
167. Bahramisharif, A.; Mazaheri, A.; Levar, N.; Schuurman, P.R.; Figeet, M.; Denys, D. Deep Brain Stimulation Diminishes Cross-Frequency Coupling in Obsessive-Compulsive Disorder. *Biol. Psychiatry* **2016**, *80*, e57–e58. <https://doi.org/10.1016/j.biopsych.2015.05.021>.
168. Widge, A.S.; Zorowitz, S.; Link, K.; Miller, E.K.; Deckersbach, T.; Eskandar, E.N.; Dougherty, D.D. Ventral Capsule/Ventral Striatum Deep Brain Stimulation Does Not Consistently Diminish Occipital Cross-Frequency Coupling. *Biol. Psychiatry* **2016**, *80*, e59–e60. <https://doi.org/10.1016/j.biopsych.2015.10.029>.
169. Kamimura, H.A.; Wang, S.; Chen, H.; Wang, Q.; Aurup, C.; Acosta, C.; Carneiro, A.A.; Konofagou, E.E. Focused ultrasound neuromodulation of cortical and subcortical brain structures using 1.9 MHz. *Med. Phys.* **2016**, *43*, 5730. <https://doi.org/10.1118/1.4963208>.
170. Downs, M.E.; Lee, S.A.; Yang, G.; Kim, S.; Wang, Q.; Konofagou, E.E. Non-invasive peripheral nerve stimulation via focused ultrasound in vivo. *Phys. Med. Biol.* **2018**, *63*, 035011. <https://doi.org/10.1088/1361-6560/aa9fc2>.
171. Rodenkirch, C.; Wang, Q. Rapid and transient enhancement of thalamic information transmission induced by vagus nerve stimulation. *J. Neural Eng.* **2020**, *17*, 026027. <https://doi.org/10.1088/1741-2552/ab6b84>.
172. Legon, W.; Sato, T.F.; Opitz, A.; Mueller, J.; Barbour, A.; Williams, A.; Tyler, W.J. Transcranial focused ultrasound modulates the activity of primary somatosensory cortex in humans. *Nat. Neurosci.* **2014**, *17*, 322–329. <https://doi.org/10.1038/nn.3620>.

173. McIntyre, C.C.; Grill, W.M. Extracellular Stimulation of Central Neurons: Influence of Stimulus Waveform and Frequency on Neuronal Output. *J. Neurophysiol.* **2002**, *88*, 1592–1604. <https://doi.org/10.1152/jn.2002.88.4.1592>.
174. Wang, Q.; Millard, D.C.; Zheng, H.J.V.; Stanley, G.B. Voltage-sensitive dye imaging reveals improved topographic activation of cortex in response to manipulation of thalamic microstimulation parameters. *J. Neural Eng.* **2012**, *9*, 026008.
175. Bari, B.A.; Ollerenshaw, D.R.; Millard, D.C.; Wang, Q.; Stanley, G.B. Behavioral and electrophysiological effects of cortical microstimulation parameters. *PLoS ONE* **2013**, *8*, e82170. <https://doi.org/10.1371/journal.pone.0082170>.
176. Millard, D.C.; Wang, Q.; Gollnick, C.A.; Stanley, G.B. System identification of the nonlinear dynamics in the thalamocortical circuit in response to patterned thalamic microstimulation in vivo. *J. Neural Eng.* **2013**, *10*, 066011.
177. Mickle, A.D.; Gereau, R.W.I. A bright future? Optogenetics in the periphery for pain research and therapy. *Pain* **2018**, *159*, S65–S73. <https://doi.org/10.1097/j.pain.0000000000001329>.
178. Boggio, P.S.; Rigonatti, S.P.; Ribeiro, R.B.; Myczkowski, M.L.; Nitsche, M.A.; Pascual-Leone, A.; Fregni, F. A randomized, double-blind clinical trial on the efficacy of cortical direct current stimulation for the treatment of major depression. *Int. J. Neuropsychopharmacol.* **2008**, *11*, 249–254. <https://doi.org/10.1017/s1461145707007833>.
179. Zangen, A.; Moshe, H.; Martinez, D.; Barnea-Ygaël, N.; Vapnik, T.; Bystritsky, A.; Duffy, W.; Toder, D.; Casuto, L.; Grosz, M.L.; et al. Repetitive transcranial magnetic stimulation for smoking cessation: A pivotal multicenter double-blind randomized controlled trial. *World Psychiatry* **2021**, *20*, 397–404. <https://doi.org/10.1002/wps.20905>.
180. Sadleir, R.; Vannorsdall, T.; Schretlen, D.; Gordon, B. Target Optimization in Transcranial Direct Current Stimulation. *Front. Psychiatry* **2012**, *3*, 90. <https://doi.org/10.3389/fpsy.2012.00090>.
181. Khadka, N.; Truong, D.Q.; Bikson, M. Principles of Within Electrode Current Steering1. *J. Med. Devices* **2015**, *9*, 020947. <https://doi.org/10.1115/1.4030126>.
182. Slater, C.; Wang, Q. Alzheimer’s disease: An evolving understanding of noradrenergic involvement and the promising future of electroceutical therapies. *Clin. Transl. Med.* **2021**, *11*, e397. <https://doi.org/10.1002/ctm2.397>.
183. Sharon, O.; Fahoum, F.; Nir, Y. Transcutaneous Vagus Nerve Stimulation in Humans Induces Pupil Dilation and Attenuates Alpha Oscillations. *J. Neurosci. Off. J. Soc. Neurosci.* **2021**, *41*, 320–330. <https://doi.org/10.1523/JNEUROSCI.1361-20.2020>.
184. Turi, Z.; Mittner, M.; Lehr, A.; Bürger, H.; Antal, A.; Paulus, W. θ - γ Cross-Frequency Transcranial Alternating Current Stimulation over the Trough Impairs Cognitive Control. *eNeuro* **2020**, *7*, ENEURO.0126-0120.2020. <https://doi.org/10.1523/ENEURO.0126-20.2020>.
185. Polanía, R.; Nitsche, M.A.; Korman, C.; Batsikadze, G.; Paulus, W. The Importance of Timing in Segregated Theta Phase-Coupling for Cognitive Performance. *Curr. Biol.* **2012**, *22*, 1314–1318. <https://doi.org/10.1016/j.cub.2012.05.021>.
186. Riddle, J.; McFerren, A.; Frohlich, F. Causal role of cross-frequency coupling in distinct components of cognitive control. *Prog. Neurobiol.* **2021**, *202*, 102033. <https://doi.org/10.1016/j.pneurobio.2021.102033>.
187. Yazdan-Shahmorad, A.; Silversmith, D.B.; Sabes, P.N. Novel techniques for large-scale manipulations of cortical networks in non-human primates. In Proceedings of the 2018 40th Annual International Conference of the IEEE Engineering in Medicine and Biology Society (EMBC), Honolulu, HI, USA, 18–21 July 2018; Volume 2018, pp. 5479–5482. <https://doi.org/10.1109/EMBC.2018.8513668>.
188. Widge, A.S.; Miller, E.K. Targeting Cognition and Networks Through Neural Oscillations: Next-Generation Clinical Brain Stimulation. *JAMA Psychiatry* **2019**, *76*, 671–672. <https://doi.org/10.1001/jamapsychiatry.2019.0740>.

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